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## The measurement of the rate of transport of solutes in both directions across the aqueous-nonaqueous liquid interface and its significance to membrane permeability

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A technique for measuring the partition coefficients,  $K_{pc}$ , and the rate of movement of solutes across an aqueous-nonaqueous interface is described. Using it, measurements of the rate constants for the transport of water and the lower alcohols in both directions across the water-octanol interface were made. The effect of unstirred layers was found to be unimportant in the present case. The partition coefficients of water and the alcohols, calculated as the ratio of the rate constants, showed good agreement with those measured directly. For small molecules, such as the lower alcohols, the permeability constants of natural membranes do not appear to be determined either by  $K_{pc}$  or the rate constants, probably due to the presence of pores. Since the resistance offered to water transport across the interface is as high as that of any artificial lipid bilayer for which measurements have been reported, it appears that the main barrier to permeation in these structures is not diffusion within the membrane, but transport across the aqueous-nonaqueous interface.

### Introduction

When two immiscible liquids are in contact, movement of any solutes which are soluble in both phases will occur across the interface separating them. The rate of solute transfer out of each phase is proportional to its concentration in that phase. Thus for two liquids (labelled 'o' for the organic and 'w' for the water phase) the rates will be  $k_{ow}C_o$  from the organic to water phase and  $k_{wo}C_w$  in the reverse direction, where  $C_o$  and  $C_w$  are the respective concentrations and  $k_{ow}$  and  $k_{wo}$  are the proportionality, or transfer rate constants. Transfer will continue as long as the solute is present and the phases are in contact; however, net transfer between the phases will take place only until the rates of movement in both directions are equal. Such equilibrium will be attained only after the elapse of an infinite time, and if the concentrations at this point are  $C_o^\infty$  and  $C_w^\infty$ , then since  $k_{ow}C_o^\infty =$

$k_{wo}C_w^\infty$  we can define a term known as the partition coefficient,  $K_{pc}$ , as  $K_{pc} = C_o^\infty / C_w^\infty = k_{wo} / k_{ow}$ .

The evaluation of  $K_{pc}$  for a given solute-solvent system is relatively easy and as a consequence many  $K_{pc}$  values appear in the literature (see for example Ref. 1). In contrast, the measurements of individual rate constants require much more sophisticated methods, with the result that relatively few such measurements have been reported. This is true in spite of the obvious importance of the dynamic aspects of interfacial transport, particularly in such fields of study as biological membrane permeability. What is particularly surprising is that so little is known concerning the effect on the aqueous/organic-solvent interfacial transport of substances adsorbed at the interface, and it was with the view of obtaining such knowledge that the present studies were initiated. This report describes the experimental techniques to be employed and illustrates and evaluates them using a

relatively simple system involving the movement of water and the lower alcohols between the water and octanol phases. Octanol was chosen as the organic phase since its solvent properties have been found to resemble those of biological systems more closely than other solvents.

## Experimental

Three basic experimental systems have been employed in interfacial transport studies: layers, jets and falling drops [2]. Most organic liquids have a density of less than one and can be layered over water. In this configuration the rates of passage of solute from one phase to the other can be measured simply by quantitative analysis of the solutes in samples taken from either layer as a function of time. The problem with this simple system is that solute concentration gradients, which form on each side of the interface, develop a resistance to solute diffusion with time which can rapidly exceed that of transfer across the interface. Even with stirring there remains a layer of liquid near the interface which is undisturbed. The thickness of this unstirred layer is reduced as the stirring rate increases. At the same time, however, the onset of turbulence with rapid stirring distorts the interface to the point where its area becomes uncertain. This problem is avoided in the rotating diffusion cell [3], in which the organic liquid is confined to a filter paper and is therefore undisturbed by rapid stirring. In this method, rate measurements made at different stirring speeds are extrapolated to infinite speed, at which the resistance of the unstirred layer can be assumed to approach zero.

In the second experimental system, one phase moves through the other in the form of a well-defined jet, the surface area of which can be readily measured. Unstirred layers in such a system will, of course, be negligible. However, only very rapid transport rates can be measured by this method, which is therefore inappropriate for the present study.

In the third system, one phase is stationary while the second passes through it in the form of a drop rising (or falling) under gravity. As the drop passes through the continuous phase, liquid is swept from its leading to its trailing surface, caus-

ing rapid internal stirring and reducing the drag which the continuous phase would otherwise produce on the drop. Thus the liquid on both sides of the interface is constantly renewed and unstirred layers have little chance to form [2,4]. At the same time, since the drop is a sphere, its surface area is well defined and can be determined accurately. It is the appealing simplicity of this system which prompted its use in the present study.

**Materials.** Octan-1-ol (99%) was obtained from Aldrich and used directly from the bottle after saturation with water; methanol was redistilled reagent-grade; ethanol was absolute from Consolidated Alcohols Ltd. Toronto (pharmaceutical grade); propan-1-ol was obtained from Matheson, Coleman and Bell; butan-1-ol was HPLC-grade supplied by Fisher Scientific Co.;  $^3\text{H}$ HO was obtained from New England Nuclear with a specific activity of 25 mCi/g.

**Apparatus and procedure.** Two versions of the apparatus used to measure the rate constants and partition coefficients are shown in Fig. 1. In version A, drops of the organic phase rose through a column of the aqueous phase and were sampled at the top, while in version B the aqueous phase was dropped through a column containing the organic phase and was sampled at the bottom. Before use, each phase was saturated with the other at the temperature used for the measurements.

The measurement procedure for the movement of solute out of an octanol drop in the version shown in Fig. 1A was as follows. Syringe  $S_1$  was filled with the organic phase containing the solute and  $S_3$  was filled with the aqueous phase. Syringe  $S_2$  contained the aqueous phase which had been previously equilibrated with the solute in the organic phase. The reason for this is that drops were formed by injecting 30  $\mu\text{l}$  of the solute-containing organic phase into the capillary from  $S_1$  at the bottom of the apparatus in the position shown (D) and were then forced out of the top of the capillary by rapidly injecting 40–50  $\mu\text{l}$  of fluid from  $S_2$  into the apparatus. Since this fluid had already been equilibrated with the drop, no movement of solute occurred until the fully formed drop was ejected from the capillary. The remainder of the apparatus contained the aqueous phase which the stirring circulated in the direction shown by the arrows. To start a measurement, a

sample tube (S) consisting of a piece of melting-point capillary-glass tubing (2–2.5 cm long and about 0.1 cm i.d.) was inserted into the top of a short piece of silicon rubber tubing whose bottom end was connected to the top of the apparatus. After injecting the 30  $\mu$ l of organic phase into the capillary at the bottom of the apparatus, the meniscus at the top of the aqueous phase ( $M_1$ ) was positioned by operating  $S_3$  so that, following the injection of the drop using  $S_2$ , it would still be a few millimeters below the sample tube. A timer was started at the instant the drop was injected and the stirring rate was adjusted so that the drop rose through the wider section of the column to

point L, but was unable to enter the narrower section above it due to the higher velocity there. After the required time had elapsed, stirring was terminated, allowing the drop to continue its rise. At the instant it broke through the upper meniscus ( $M_1$ ), the timer was stopped and by manipulating  $S_3$  about 10–20  $\mu$ l of the drop contents were forced into the sample tube, which was immediately separated from the apparatus. Analysis of this liquid provided a measure of the solute concentration in the drop as a function of the time interval. Usually, 15–20 similar measurements, but with varying time intervals, were used to estimate the rate constants, as detailed below. In experi-

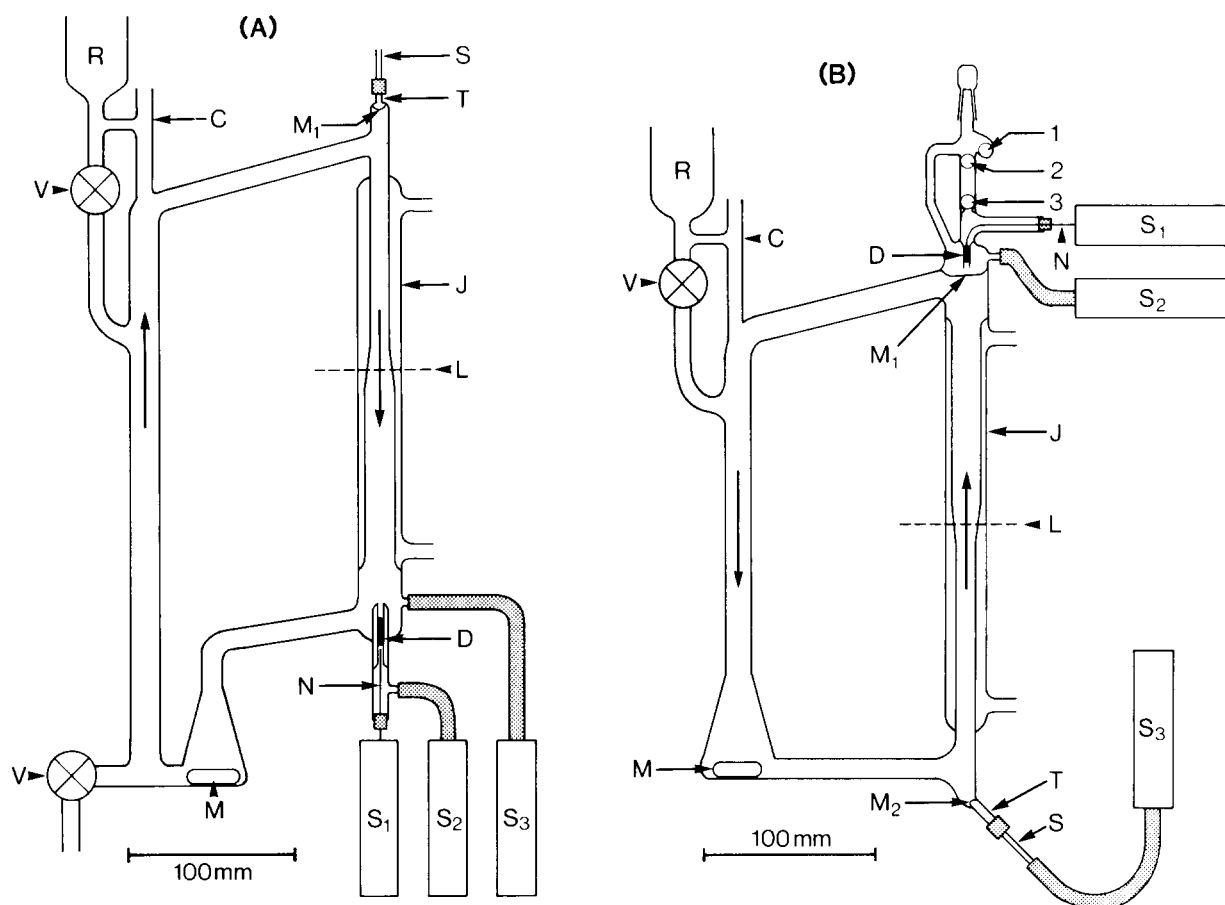


Fig. 1. (A) Rising drop apparatus, for the measurement of the rate of movement of solute between a drop of organic phase rising through a column of aqueous phase. (B) Falling drop apparatus, for the measurement of the rate of movement of solute between a drop of aqueous phase falling through a column of organic phase. C, thermometer port; D, drop-forming medium; J, water jacket; L, level at which drop is suspended by stirring; M, magnetic stirrer;  $M_1$ , upper meniscus;  $M_2$ , lower meniscus; N, syringe needle; R, reservoir; S, sample collecting tube;  $S_1$ ,  $S_2$  and  $S_3$ , 2 ml micrometer syringes; T, tube to which sample tube is attached; V, glass-Teflon valve; 1, 2 and 3, positions of steel ball.

ments in which solute was moving into the drop, the solute was present in the bulk aqueous phase but absent from the liquids in the syringes. In addition, several measurements were made after time intervals of sufficient length to ensure virtual equilibrium, and the average of these measurements was used to determine  $K_{pc}$ .

The rate of movement of solute between an aqueous phase drop and the continuous organic phase was measured using the apparatus shown in Fig. 1B. Since the drop fell through the continuous phase it was introduced at the top and collected at the bottom. In this experiment, syringe  $S_1$  contained the drop-forming solution, while  $S_2$  and  $S_3$  contained air. 30  $\mu$ l of aqueous solution were deposited in the position shown (D, Fig. 1B) with the sample tube (S) and silicon rubber tubing disconnected from tube T. A steel ball-bearing in the upper part of the apparatus was lifted by a magnet to position 1 (Fig. 1B), and by enlarging the volume of  $S_2$ , air was drawn into tube T. A sample tube was then connected to T by means of the silicon rubber tubing and  $S_3$  was connected to its other end. Adjusting  $S_3$  gave a slightly convex meniscus ( $M_2$ ) at the upper end of T. To start a measurement, the steel ball was moved to position 2, then dropped to position 3, forcing air downward and ejecting the drop which fell into the column of organic liquid. Again, the stirring was adjusted so that the drop was suspended at L for the required time, then allowed to drop further by terminating stirring. A timer which was started when the drop passed through  $M_1$  was stopped when it passed through  $M_2$ . As before, 10–20  $\mu$ l of the drop contents were drawn into the sample tube by manipulating  $S_3$ , and analyzed.

When not in use, both versions of the apparatus were filled with normal KOH. This treatment of the glass surface caused it to repel the organic phase so that drops of the latter would not adhere to it and, furthermore, aided sampling by forming more well-defined menisci.

**Analytical procedures.** Two methods were used to analyze the samples: gas chromatography for the alcohols, and scintillation counting for the  $^3\text{HHO}$ . For GC, 0.5–2  $\mu$ l samples of the liquid in the sample tube were withdrawn and injected into a Hewlett Packard 5880A series chromatograph with flame ionization detector and a Chrompack

(Middelburg, The Netherlands) CP<sup>TM</sup> Wax-57-CB fused silica capillary column. The procedure used to measure tritiated water movement comprised weighing scintillation vials containing sample tubes, removing the tubes to collect the sample, returning them to the vial and reweighing to give the sample weight by difference. Scintillation fluid was then added to the vials and the radioactive contents were assayed.

## Calculations

If  $n$  is the number of moles of solute in an octanol drop of area  $A$ , the flux of solute into the drop will be

$$dn/dt = A(k_{wo}C_w - k_{ow}C_o)$$

Due to the large volume of the water phase relative to that of the drop,  $C_w$  is a constant and only  $C_o$  is a variable. After an infinite time the system reaches equilibrium and net transport ceases. At this point,  $dn/dt = 0$  and  $C_o$  has the value  $C_o^\infty$ . Furthermore, since influx must equal efflux,  $k_{wo}C_w = k_{ow}C_o^\infty$  and therefore we can rewrite the original equation as  $dn/dt = Ak_{ow}(C_o^\infty - C_o)$  or, for a drop of volume  $V$ , letting  $C_o = n/V$ ,

$$dC_o/dt = (A/V)k_{ow}(C_o^\infty - C_o)$$

Integrating this expression between the limits  $C_o = C_o^0$  at  $t = 0$  and  $C_o$  at time  $t$ , gives

$$\ln(C_o^\infty - C_o) = \ln(C_o^\infty - C_o^0) - Ak_{ow}t/V \quad (1)$$

Measuring  $C_o$  as a function of  $t$  and plotting the results according to Eqn. 1 should provide a straight line whose slope is  $-Ak_{ow}/V$ , or for a drop whose volume is 0.030  $\text{cm}^3$ ,  $k_{ow}$  = the slope multiplied by  $-642.5 \mu\text{m}$ . The advantage of this procedure for measuring  $k_{ow}$  is that the initial concentration in the drop,  $C_o^0$ , need not be known. Thus if there is a constant error in initiating the time measurement, in which case the assumed value of  $C_o^0$  is not the true concentration of the drop-forming solution, this will not affect the measurement of  $k_{ow}$ , and one possible source of error is eliminated.

Rate measurements may be made in two ways: either the solute is initially present in the drop, but

not in the continuous phase, and its loss from the drop is measured; or it is present initially only in the continuous phase, in which case uptake by the drop is determined. When the rate of loss is measured,  $C_o^\infty = 0$  and Eqn. 1 can be rearranged to

$$\ln C_o = \ln C_o^0 - Ak_{ow}t/V \quad (2)$$

For uptake measurements, expression 1 is used but  $C_o^\infty$  must be determined. In practice, a sufficient number of measurements at different times were made to estimate  $t_{1/2}$ , the half-time of the uptake. Measurements of  $C_o$  were then made after a time interval of ten half-lives, at which point the concentration should be greater than 99.9% of the true infinite time value. This quantity (taken as  $C_o^\infty$ ) can be used to determine  $K_{pc}$ , which is equal to  $C_o^\infty$  divided by  $C_w$ , the concentration of solute in the water phase.

It should be noted that whether solute movement, as measured by the apparatus in Fig. 1A, is into or out of the octanol drop, only  $k_{ow}$  is determined according to Eqns. 1 and 2. The rate constant  $k_{wo}$  must be measured using a water drop in the apparatus depicted in Fig. 1B, and is calculated from equations derived in a similar manner to 1 and 2. These have the form

$$\ln(C_w^\infty - C_w) = \ln(C_w^\infty - C_w^0) - Ak_{wo}t/V \quad (3)$$

and

$$\ln(C_w) = \ln(C_w^0) - Ak_{wo}t/V \quad (4)$$

where  $C_w^\infty$  and  $C_w^0$  are the infinite and zero-time concentrations of solute in the water drop. Since 30  $\mu$ l drops were used in all the measurements reported here, the rate constants were found by multiplying the slopes of the lines, plotted according to Eqns. 1–4, by  $-V/A = -642.5 \mu\text{m}$ .  $C_w^\infty$  was determined in the same way as  $C_o^\infty$  and was used to estimate the partition coefficient  $K_{pc} = C_o/C_w^\infty$ .

In measuring the tritiated water transfer, the exchange of tritium atoms between water and octanol,  $\text{OcOH} + {}^3\text{HHO} \rightleftharpoons \text{OcO}^3\text{H} + \text{H}_2\text{O}$  (where Oc represents the octyl radical), must be considered. This reaction has been shown [5] to occur with a half-time of  $10^{-2}$ – $10^{-3}$  s and may therefore be considered to be instantaneous for the

present purposes. Thus, when tritiated water enters an octanol drop, some of the tritium will exchange with octanol, reducing the radioactivity of the water which has entered the drop. The net movement of  ${}^3\text{HHO}$  will continue into the drop until its activity there equals that in the water phase. This will require that more  ${}^3\text{HHO}$  enter the drop than is required to raise the activity of the water in the drop alone, since that of the octanol must also rise to its equilibrium value. Since there is 1 mol of exchangeable hydrogen atom per mol of octanol and 2 mol of exchangeable hydrogen atoms per mol of water, the radioactive dpm of 9 g water should be equal to that of 130.23 g octanol at equilibrium. Defining the activities as  $a_w$  = dpm/g water in the drop and  $a_o$  = dpm/g octanol in the drop, then  $a_o/a_w = 0.06911$  at all times. The total dpm in the drop will be the sum of that due to the water and octanol in the drop, or  $a_d w_d = a_w w_w + a_o w_o$ , where  $a_d$  is the total activity of the drop, and the weight of the drop,  $w_d$ , is the weight of the water,  $w_w$ , plus that of the octanol in the drop,  $w_o$ . Thus  $a_d/a_w = 0.9309(w_w/w_d) + 0.06911$ . Since the octanol phase was saturated with water throughout the measurements,  $w_w$  and  $w_d$  are constant and their ratio  $w_w/w_d = S$ , the solubility of water in the octanol phase in g/g. After the two phases have been in contact for an infinite length of time, the activity of the water in the drop,  $a_w^\infty$ , will equal that of the bulk water phase, and  $a_d$  will have reached its equilibrium value,  $a_d^\infty$ . Letting the ratio at this point be  $R^\infty = a_d^\infty/a_w^\infty$  (i.e., the dpm/g of the octanol phase divided by the dpm/g of the water phase at infinite time),

$$S = (R^\infty - 0.06911)/0.9309$$

To derive an expression for  ${}^3\text{HHO}$  movement equivalent to Eqn. 1, let  $n_t$  be the total number of moles of  ${}^3\text{H}$  in the drop (as both  ${}^3\text{HHO}$  and octan-1- $[{}^3\text{H}]$ ol) while  $C_w$  and  $C_o$  are the concentrations of  ${}^3\text{HHO}$  alone in the bulk and drop, respectively. If  $n_w$  = moles of  ${}^3\text{HHO}$  in the drop,  $C_o = n_w/V$  and the ratio of this to  $n_t$  will be

$$\frac{n_w}{n_t} = \frac{\text{dpm}_w}{\text{dpm}_d} = \frac{a_w w_w}{a_d w_d} = S \frac{a_w}{a_d}$$

Due to the rapidity of exchange of  $^3\text{H}$  between water and octan-1-ol, the ratio of the activity of the water in the drop to that of the drop as a whole will be constant and therefore equal to its value at infinite time. Thus  $a_d/a_w = R^\infty$  and  $n_w = (S/R^\infty)n_i$ . Next, letting  $C_i = n_i/V$  be the total concentration of  $^3\text{H}$  in the drop, and  $C_b$  that in the bulk water, (which is constant because of the large volume),

$$\frac{dC_i}{dt} = \frac{A}{V} \left( k_{wo}C_b - \left( \frac{S}{R^\infty} \right) k_{ow}C_i \right)$$

At infinite time  $dC_i/dt = 0$  and  $C_i$  reaches the value  $C_i^\infty$ . Thus

$$\frac{dC_i}{dt} = \frac{AS}{VR^\infty} k_{ow} (C_i^\infty - C_i)$$

The activity of the drop will be proportional to  $C_i$ , or  $C_i = pa_d$ , where  $p$  is a constant. In practice  $R$  is taken as the ratio of the cpm per g of drop phase divided by the cpm per g of the water phase and as this equals  $a_d/a_b$ , substituting  $pa_bR$  for  $C_i$  and  $pa_bR^\infty$  for  $C_i^\infty$  and integrating, we obtain

$$\ln(R^\infty - R) = \ln(R^\infty - R^0) - (AS/VR^\infty) k_{ow} t$$

In this case,  $k_{ow}$  is the slope of the line multiplied by the factor  $(-VR^\infty/AS)$ . This is true also for the loss of  $^3\text{HHO}$  from an octan-1-ol drop (equivalent to Eqn. 2) but not for the measurements with a water drop where the factor is, as before,  $(-V/A)$ , so that Eqns. 3 and 4 still apply here.

TABLE I

TRANSFER RATE CONSTANTS AND OCTANOL/WATER PARTITION COEFFICIENTS FOR  $^3\text{HHO}$  AND THE LOWER ALCOHOLS

Temperature was 25°C. Rate constants are in units of  $\mu\text{m}\cdot\text{s}^{-1} \pm \text{S.D.}$

Compound	$k_{ow}$ (octanol drop)		$k_{wo}$ (water drop)		Partition coefficient		
	loss	uptake	loss	uptake	octanol drop	water drop	$k_{wo}/k_{ow}$
$^3\text{HHO}$	$44.4 \pm 3.4$	$44.6 \pm 4.5$	$2.3 \pm 0.2$	$2.3 \pm 0.2$	0.0524	0.0535	0.0517
Methanol	$50.5 \pm 3.7$	$45.0 \pm 3.0$	$11.5 \pm 1.0$	$10.1 \pm 0.9$	0.245	0.203	0.227
Ethanol	$43.9 \pm 1.5$	$40.1 \pm 1.7$	$20.1 \pm 1.3$	$20.9 \pm 2.8$	0.493	0.481	0.488
<i>n</i> -Propanol	$25.6 \pm 0.5$	$28.3 \pm 2.3$	$56.4 \pm 2.7$	$52.3 \pm 2.7$	2.21	1.94	2.02
<i>n</i> -Butanol	$11.5 \pm 0.3$	$10.8 \pm 0.2$	$65 \pm 17$	$73 \pm 20$	7.12	7.03	6.2

TABLE II

TRANSFER RATE CONSTANTS AND PARTITION COEFFICIENT FOR *n*-BUTANOL BETWEEN WATER AND OCTANOL AS A FUNCTION OF TEMPERATURE

Both  $k_{ow}$  and  $K_{pc}$  were measured as uptake of butanol into an octanol drop from a water solution of 10 mM. The  $k_{wo}$  values (in brackets) were calculated as  $k_{wo} = k_{ow}K_{pc}$ .

$T$ (°C)	$k_{ow}$	$K_{pc}$	$k_{wo}$
15	$8.8 \pm 0.5$	6.07	(53.4)
20	$10.6 \pm 0.5$	6.01	(63.7)
25	$10.0 \pm 0.4$	6.46	(64.6)
30	$14.0 \pm 0.5$	7.02	(98.3)
35	$15.4 \pm 0.5$	7.67	(118)

## Results

Table I lists the rate constants and partition coefficients of all compounds at 25°C. Similar determinations made at various temperatures are reported for butanol in Table II and water in Table III. Arrhenius plots of these data give activation energies for the transport of butanol which are  $4.9 \pm 1.0 \text{ kcal}\cdot\text{mol}^{-1}$  in the direction octanol to water and  $2.2 \pm 0.4 \text{ kcal}\cdot\text{mol}^{-1}$  in the opposite direction. The corresponding values for water are  $6.9 \pm 0.4$  and  $6.6 \pm 0.7 \text{ kcal}\cdot\text{mol}^{-1}$ . The density of water-saturated octanol was found to be  $0.833 \text{ g/cm}^3$  at 25°C.

According to equations in the previous section, letting  $C_i = C_i^\infty$  (and  $dC_i/dt = 0$ ) and substituting  $pa_w^\infty$  for  $C_b$ , and  $pa_d^\infty$  for  $C_i^\infty$ , it can be shown that  $k_{wo}/k_{ow} = S$ . Thus the solubility of water in the octanol phase can be obtained in three ways:

TABLE III

TRANSFER RATE CONSTANTS AND THE SOLUBILITY IN OCTANOL FOR WATER AS A FUNCTION OF TEMPERATURE

All constants were measured as uptake into a drop with the exception of those marked by an asterisk, which were measured as loss. Units are  $\mu\text{m}\cdot\text{s}^{-1}$  for the rate constants and g water per g water-saturated-octanol for the water solubility.

$T$ ( $^{\circ}\text{C}$ )	Octanol drop		Water drop		$k_{\text{wo}}/k_{\text{ow}}$
	$k_{\text{ow}}$	solubility	$k_{\text{wo}}$	solubility	
15	$29.2 \pm 2.9$	0.0452	$1.44 \pm 0.15$	0.0504	0.0493
20	$38.5 \pm 3.1$	0.0483	$2.04 \pm 0.12$	0.0528	0.0530
25	$44.6 \pm 4.9$	0.0524	$2.28 \pm 0.20$	0.0535	0.0511
25	$44.4 \pm 3.4$ *		$2.31 \pm 0.19$ *		0.0520
30	$51.5 \pm 4.2$	0.0509	$2.74 \pm 0.35$	0.0594	0.0532
35	$68.7 \pm 4.0$	0.0544	$3.53 \pm 0.21$	0.0560	0.0514
40	$77.3 \pm 6.1$	0.0500	$3.58 \pm 0.20$	0.0557	0.0463

by the direct measurement of equilibrium concentration in an octanol drop; in the same way in a water drop; and from the ratio of the rate constants. These results may be compared directly in Tables I and III. Averaging these results at  $25^{\circ}\text{C}$  (Table III) the solubility of water is found to be  $0.052 \pm 0.002$  g per g octanol phase.

## Discussion

### Effect of unstirred layers

The validity of the results depends on the magnitude of the effects of the unstirred layers. According to Dainty [6], if  $\delta_o$  and  $\delta_w$  are the thicknesses of these layers in octanol and water, and  $D_o$  and  $D_w$  are the diffusion coefficients in

the same media, the apparent rate constants  $k'_{\text{ow}}$  and  $k'_{\text{wo}}$  will be given by the expressions

$$\frac{1}{k'_{\text{ow}}} = \frac{1}{k_{\text{ow}}} + \frac{\delta_o}{D_o} + \frac{\delta_w}{D_w} \quad \text{and} \quad \frac{1}{k'_{\text{wo}}} = \frac{1}{k_{\text{wo}}} + \frac{\delta_o}{D_o} + \frac{\delta_w}{D_w}$$

Since the resistance to diffusion can be taken as the reciprocal of the rate constant, then if the resistances to passage of the solute across the interface in the two directions are  $R_{\text{ow}} = 1/k_{\text{ow}}$  and  $R_{\text{wo}} = 1/k_{\text{wo}}$ , and  $R_o = \delta_o/D_o$  and  $R_w = \delta_w/D_w$  are the resistances of the unstirred layers, the apparent resistances will be  $R'_{\text{ow}} = R_{\text{ow}} + R_u$  and  $R'_{\text{wo}} = R_{\text{wo}} + R_u$ , where  $R_u = R_o + R_w$ . ( $\delta_o$  is assumed to be the same in the drop as in the continuous medium. The same assumption is made about  $\delta_w$ .) The apparent partition coefficient as derived from the rate constants will therefore be

$$K'_{\text{pc}} = \frac{k'_{\text{wo}}}{k'_{\text{ow}}} = \frac{R'_{\text{ow}}}{R'_{\text{wo}}} = \frac{R_{\text{ow}} + R_u}{R_{\text{wo}} + R_u} = \frac{K_{\text{pc}} + r}{1 + r}$$

whereas the true constant  $K_{\text{pc}} = R_{\text{ow}}/R_{\text{wo}}$ . Here  $r$  is the ratio of the unstirred layer resistance to that of the interface ( $r = R_u/R_{\text{wo}}$ ). This ratio may also be written as  $r = R_u k_{\text{wo}}$ , which tells us that when  $\delta$  and hence  $R_u$  are small,  $K'_{\text{pc}} \approx K_{\text{pc}}$ , but as the layer thickness, or the constant  $k_{\text{wo}}$ , increases,  $K'_{\text{pc}}$  approaches one. This effect is apparent in Table I where, for compounds having lower values of  $k_{\text{wo}}$ , there is good agreement between the values for  $K_{\text{pc}}$  measured directly as a ratio of concentrations in the two phases and those measured indirectly as

TABLE IV

COMPARISON OF THE MEASURED AND CALCULATED VALUES OF MEMBRANE PERMEABILITY

The partition coefficients,  $K_{\text{pc}}$ , are averages from the present work. The calculated permeability constants  $= k_{\text{wo}}/2$ , while the measured constants are for plant and animal cells. All rate constants are in  $\mu\text{m}\cdot\text{s}^{-1}$ .

Compound	$K_{\text{pc}}$	Calculated	Measured values (Ref.)		
			8	9	10
Water	0.052	1.2	91.5	—	25
Methanol	0.225	5.4	1.14	3.3	5.7
Ethanol	0.487	10	0.88	2.4	5.5
<i>n</i> -Propanol	2.06	27	0.64	2.6	7.2
<i>n</i> -Butanol	7.08	35	0.41	2.5	—

the ratio  $k_{wo}/k_{ow}$ . For butanol, which has the highest value of  $k_{wo}$  reported here, however, the latter value appears to be lower. Thus we may conclude that for 30  $\mu\text{l}$  drops of these two liquids, internal stirring is sufficient to eliminate unstirred layer effects, provided the interfacial transfer rate constants do not exceed 60  $\mu\text{m} \cdot \text{s}^{-1}$ .

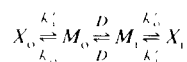
The importance of stirring within the drop can be further illustrated by comparing the rates measured here against those expected in the absence of stirring. The latter may be obtained from the equation given by Jost [7] which provides the average concentration of a diffusing solute,  $\bar{C}$ , in a sphere of radius  $r_o$ , as a function of time,  $t$ . If  $C^0$  is the initial and  $C^\infty$  the final concentration, then for  $t$  which is sufficiently large:

$$\frac{\bar{C} - C^\infty}{C^0 - C^\infty} \approx \frac{6}{\pi^2} \exp \left[ -\frac{\pi^2 D}{r_o^2} t \right]$$

Letting  $\bar{C} = 0.5C^\infty$ ,  $C^0 = 0$  (or conversely  $\bar{C} = 0.5C^0$  and  $C^\infty = 0$ ) and  $D = 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ , the half-time for diffusion in a 30  $\mu\text{l}$  sphere (i.e., in a drop with no stirring) should be 73.6 s. In contrast, half-times less than 5 s were found for the movement by butanol out of a 30  $\mu\text{l}$  water drop, showing that stirring is indeed active in the drops used here.

#### *Significance of the measurements to membrane transport*

There are three steps involved in the passive transport of a solute ( $X$ ) through the lipid bilayers of the plasmalemma:



Starting at a concentration  $X_o$  outside the cell, the solute moves across the outer membrane interface, at a rate determined by the constant  $k_i$ , to establish a concentration  $M_o$  just inside the membrane. Diffusion through the membrane next takes place at a velocity dependent on the diffusion coefficient,  $D$ , as a result of which it arrives at the opposite interface, where its concentration is  $M_i$ . Finally, movement occurs through the inner membrane interface to the cell interior (rate constant  $k'_i$ ), where its concentration is  $X_i$ . The rate of net inward transport across the outer membrane

surface will be the rate into the membrane,  $Ak_i X_o$ , minus that out,  $Ak_o M_o$ . Similarly, for the inner surface the rate will be  $A(k'_i M_i - k'_i X_i)$ . Diffusion is usually assumed to obey Fick's law from which the inward rate of diffusion can be shown to approximate  $(AD/\delta)(M_o - M_i)$ , where  $\delta$  is the membrane thickness. Under steady-state conditions, all these rates must be equal so that if  $n$  is the number of moles of solute in the cell

$$\frac{dn}{dt} = \frac{A(D/\delta)(k'_o k_i X_o - k_o k'_i X_i)}{k_o k'_o + (D/\delta)(k_o + k'_o)}$$

At infinite time,  $dn/dt = 0$  and since this type of transport is passive,  $X_o$  will equal  $X_i$  so that  $k'_o k_i$  must equal  $k_o k'_i$  and the rate expression becomes

$$\frac{dn}{dt} = \frac{A(D/\delta)k'_i k_i (X_o - X_i)}{k_o k'_o + (D/\delta)(k_o + k'_o)}$$

Finally, if  $k'_o = k_o$ , as would be the case for a symmetrical bilayer

$$\frac{dn}{dt} = \frac{Ak_i(D/\delta)}{k_o + 2(D/\delta)} (X_o - X_i) = AP(X_o - X_i)$$

with  $P = k_i(D/\delta)/(k_o + 2(D/\delta))$  being the permeability constant. If the slow step is assumed to be diffusion within the membrane, i.e. if  $k_o > D/\delta$ , then  $P \approx K_{pc} D/\delta$ , where  $K_{pc} = k_i/k_o$  is the partition coefficient of the solute between the water and the membrane lipids. This assumption is usually made, since it is found in general that for a series of compounds,  $P$  is proportional to  $K_{pc}$ . In some cases, notably with the series of compounds examined here, this simple relation does not hold and it is worth considering the alternative assumption, that transport across the interface is the slow process, or that  $D/\delta \gg k_o$ , making  $P \approx k_i/2$ . Assuming that the solvent properties of octanol closely approximate those of the membrane lipids,  $k_o$  and  $k_i$  may be replaced by  $k_{ow}$  and  $k_{wo}$ , allowing an estimate of the permeability constants for water and the lower alcohols to be made for this case. These are listed in Table IV, together with their partition coefficients, and are compared with three sets of experimental data [8–10]. Here it can be seen that while  $K_{pc}$  and the calculated values of the permeability constant increase with molecular weight, the measured permeabilities ap-



pear to be roughly constant (except for water, of course, which is anomalously high). This, coupled with the fact that the calculated values of  $P$  (again excluding water) tend to be higher than the measured values, appears to argue against interfacial transport as the slow step in passive membrane transport.

The high value for the permeability of natural membranes to water probably reflects the presence of pores [11], which might also account for the greater-than-expected permeance of smaller molecules such as methanol [9]. Lipid bilayers, on the other hand, are about an order of magnitude less permeable to water than natural membranes, having values for  $P$  which range from 1 to  $\approx 50 \mu\text{m} \cdot \text{s}^{-1}$  [11]. Since the calculated value for  $P$  found here is at the lower end of this range, interfacial transport must be the rate-limiting process for water movement in artificial bilayers, since if diffusion were slower the measured  $P$  would have to have an even lower value than the calculated value in Table IV. In short, the resistance at the aqueous-nonaqueous interface appears to account fully for the total resistance to water movement through artificial bilayers. It would be interesting to know if the same were true for the lower alcohols. Unfortunately, such a comparison cannot be made at the present time, since the values of the permeability constants of these compounds in artificial membranes have not been reported.

The activation energy for water transport across the water-octanol interface is also at the lower end of the range found for lipid bilayers at or above their transition temperatures ( $\approx 6\text{--}15 \text{ kcal} \cdot \text{mol}^{-1}$  [11,12]). Here again, interfacial transport could account for this value, but in this case the comparison is less convincing.

The higher alcohols are well-known surface-active agents and as a consequence a high degree of orientation of octanol molecules at the water interface is to be expected. The effect on transport rates produced by such orientation and packing, not only of solvent but also of other molecules (particularly lipids), is of considerable interest, especially in relation to natural membrane permeability. Such a study, using the methods described here, is presently under way.

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### References

- 1 Leo, A., Hansch, C. and Elkins, D. (1971) *Chem. Rev.* 71, 525–616
- 2 Davies, J.T. and Rideal, E.K. (1963) *Interfacial Phenomena*, Academic Press, New York
- 3 Albery, W.F., Burke, J.F., Littler, E.B. and Hadgraft, J. (1976) *J. Chem. Soc. Faraday Trans.* 72, 1618–1626
- 4 Horton, T.J., Fritsch, T.R. and Kinter, R.C. (1965) *Can. J. Chem. Eng.* 43, 143–146
- 5 Terada, H., Sherakawa, K., Kametani, F. and Yosikawa, K. (1983) *Biochim. Biophys. Acta* 725, 254–260
- 6 Dainty, J. (1963) in *Advances in Botanical Research* (Preston, R.D., ed.), Vol. 1, pp. 279–326. Academic press, New York
- 7 Jost, W. (1952) *Diffusion in Solids, Liquids and Gases*, pp. 45–46, Academic Press, Inc., New York
- 8 Collander, R. (1954) *Physiol. Plant.* 7, 420–455
- 9 Naccache, P. and Sha'afi, R.I. (1973) *J. Gen. Physiol.* 62, 714–736
- 10 Steudle, E. and Tyerman, S.D. (1983) *J. Membrane Biol.* 75, 85–96
- 11 Fettiplace, R. and Haydon, D.A. (1980) *Physiol. Rev.* 60, 510–550
- 12 Inoue, T., Kamaya, H. and Ueda, I. (1985) *Biochim. Biophys. Acta* 812, 393–401